

CHEMICAL MODIFICATION AND
ANTITUMOR ACTIVITY OF
HERBIMYCIN A.
8,9-EPOXIDE, 7,9-CYCLIC CARBAMATE,
AND 17 OR 19-AMINO DERIVATIVES

SATOSHI ŌMURA, KATSUJI MIYANO,
AKIRA NAKAGAWA, HIROSHI SANO,
KANKI KOMIYAMA and IWAO UMEZAWA

School of Pharmaceutical Sciences,
Kitasato University and The Kitasato Institute,
Minato-ku, Tokyo 108, Japan

KIYOSHI SHIBATA and
SADAYOSHI SATSUMABAYASHI

Nippon Dental University,
Chiyoda-ku, Tokyo 102, Japan

(Received for publication May 15, 1984)

Herbimycin A¹⁾ (**1**), a new ansamycin antibiotic isolated from the culture broth of *Streptomyces hygroscopicus* AM-3672, shows herbicidal, anti-tabacco mosaic virus and antitumor²⁾ activities. This compound has been shown to be a member of the benzoquinone subfamily of the ansamycins.^{3,4)} Geldanamycin^{5,6)} and macbecin,⁷⁻⁹⁾ which are also antitumor ansamycin antibiotics, have been reported to have the structures similar to **1**. Chemical modification of the amide group and the benzoquinone moiety in geldanamycin has been attempted and some derivatives were found to exhibit higher antitumor activity than the mother compound.¹⁰⁾ However, there has been no presentation of the chemical modification of the ansa-chain moiety to date.

In this paper, we describe the synthesis and antitumor activities of 8,9-epoxyherbimycin A (**2**), herbimycin A-7,9-cyclic carbamate (**3**) and the 17 or 19-amino substituted derivatives of **1**, **2** and **3**.

Treatment of **1** with *m*-chloroperbenzoic acid (1.2 equiv) in CHCl₃ gave 8,9-epoxyherbimycin A (**2**); MS *m/z* 590 (M, C₃₀H₄₂N₂O₁₀), TLC silica gel 60 F₂₅₄ plates (0.2 mm thick, Merck) Rf 0.51 (C₆H₆ - EtOAc, 1:1), [α]_D²⁵ +126° (c 0.5, CHCl₃), UV λ_{max}^{MeOH} nm (ε) 272 (23,900), IR ν_{max} (CHCl₃) cm⁻¹ 790 and 750 (assignable to the epoxy group), in 65% yield. The ¹H NMR spectrum (100 MHz CDCl₃) of **2** showed a doublet signal (*J*=

9.0 Hz) assignable to the epoxy methine proton (H-9) at δ 2.96, disappearance of a 9-olefinic proton in **1**, and up-field shifts of H-7 and 8-CH₃ proton signals to δ 4.17 and 1.33, respectively. Furthermore, the ¹³C NMR spectrum (25.1 MHz, CDCl₃) showed the disappearance of 8 and 9-olefinic carbon resonances and the appearance of 8 and 9-epoxy carbons newly formed at δ 60.2 and 67.1, respectively. On treatment of **2** with boron trifluoride etherate (3 equiv) in benzene at room temperature, herbimycin A-7,9-cyclic carbamate (**3**); MS *m/z* 590 (M, C₃₀H₄₂N₂O₁₀), TLC Rf 0.76 (C₆H₆ - EtOAc, 1:1), [α]_D²⁵ +135° (c 0.5, CHCl₃), UV λ_{max}^{MeOH} nm (ε) 274 (35,300), was obtained in 59% yield through an S_N2 attack of the 9-carbamoyl group and epoxide ring opening. The structure of **3** was confirmed from the ¹H and ¹³C NMR spectral data; δ_H 4.02 (1H, d, *J*=9.0 Hz, OCONH), 3.76 (1H, dd, *J*=5.0 and 9.0 Hz, H-9), δ_C 86.8 (C-7), 86.7 (C-8), 56.9 (C-9).

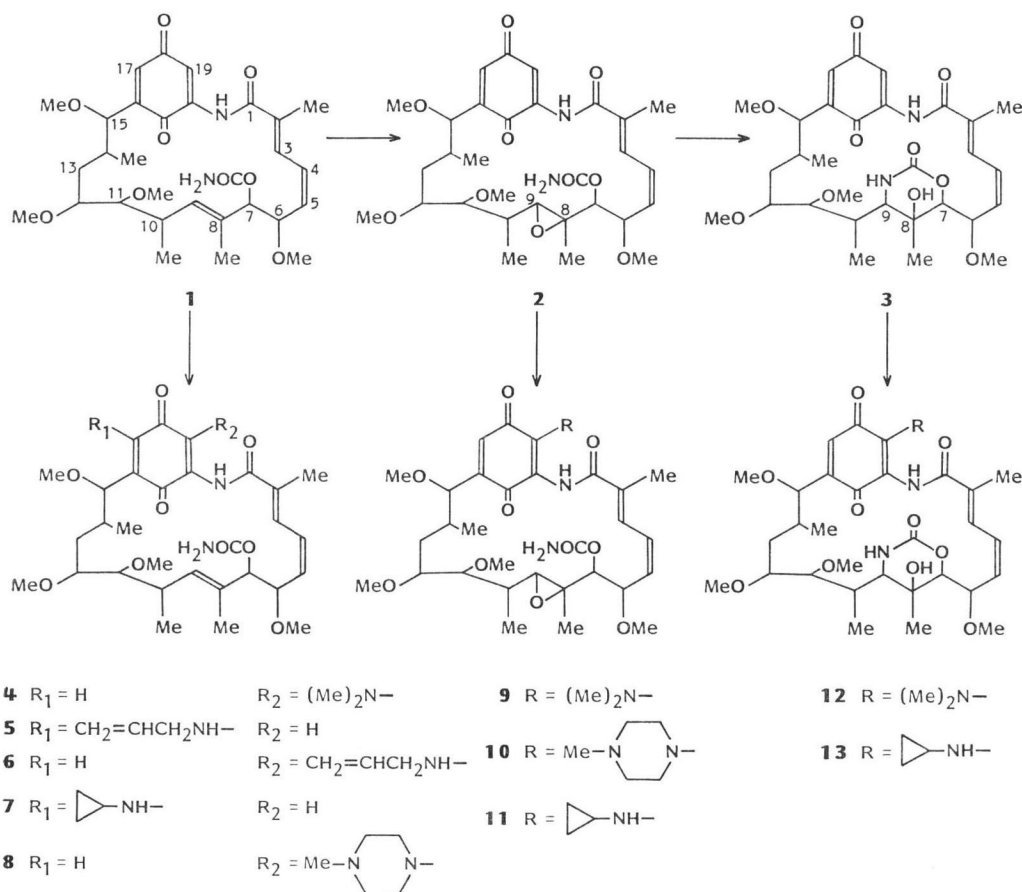
Treatment of **1**, **2** and **3** with substituted amines such as dimethylamine, allylamine, cyclopropylamine and methylpiperazine in benzene afforded the corresponding 17 or 19-substituted derivatives (**4**~**13**), respectively. This can be explained by MICHAEL addition of the nucleophile at position 17 or 19 followed by subsequent air oxidation of the resulting intermediate dihydroquinone. The structure of the amino derivatives were confirmed by the following ¹H and ¹³C NMR spectral data.

19-Dimethylaminoherbimycin A (**4**): MS *m/z* 617 (M, C₃₂H₄₇N₃O₉); TLC Rf 0.28 (C₆H₆ - EtOAc, 1:1); [α]_D²⁵ -61.5° (c 0.5, CHCl₃); UV λ_{max}^{MeOH} nm (ε) 259 (20,600); ¹H NMR (CDCl₃) δ 6.40 (1H, d, *J*=2.0 Hz, H-17), 4.34 (1H, br s, H-15), 3.17 (6H, s, N(CH₃)₂); ¹³C NMR (CDCl₃) δ 133.6 (C-17), 118.8 (C-19), 42.4 (N(CH₃)₂).

17-Allylaminoherbimycin A (**5**): MS *m/z* 629 (M, C₃₃H₄₇N₃O₉); TLC Rf 0.60 (C₆H₆ - EtOAc, 1:1); [α]_D²⁵ -150° (c 0.2, CHCl₃); UV λ_{max}^{MeOH} nm (ε) 247 (12,500) and 338 (11,700); ¹H NMR (CDCl₃) δ 7.64 (1H, m, NH), 6.96 (1H, d, *J*=2.0 Hz, H-19), 4.48 (1H, s, H-15); ¹³C NMR (CDCl₃) δ 116.9 (CH₂=CH), 109.1 (C-19), 108.2 (CH₂=CH), 48.3 (NHCH₂).

19-Allylaminoherbimycin A (**6**): MS *m/z* 629 (M, C₃₃H₄₇N₃O₉); TLC Rf 0.45 (C₆H₆ - EtOAc, 1:1); [α]_D²⁵ -124° (c 0.1, CHCl₃); UV λ_{max}^{MeOH} nm (ε) 247 (13,100) and 335 (9,000); ¹H NMR (CDCl₃) δ 7.45 (1H, m, NH), 6.48 (1H, d, *J*=1.8

Scheme 1.



Hz, H-17), 4.46 (1H, br s, H-15).

17-Cyclopropylaminoherbimycin A (7): MS m/z 629 (M, $\text{C}_{33}\text{H}_{47}\text{N}_3\text{O}_9$); TLC Rf 0.60 (C_6H_6 - EtOAc, 1:1); $[\alpha]_{\text{D}}^{25} -138^\circ$ (c 0.2, CHCl_3); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ) 245 (12,000); ^1H NMR (CDCl_3) δ 7.60 (1H, br d, NH), 6.48 (1H, d, $J=2.0$ Hz, H-17), 4.48 (1H, br s, H-15).

19-Methylpiperazinoherbimycin A (8): MS m/z 672 (M, $\text{C}_{35}\text{H}_{52}\text{N}_4\text{O}_9$); TLC Rf 0.63 (CHCl_3 - MeOH, 5:1); $[\alpha]_{\text{D}}^{25} -101^\circ$ (c 0.1, CHCl_3); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ) 257 (21,300); ^1H NMR (CDCl_3) δ 6.52 (1H, d, $J=1.8$ Hz, H-17), 4.39 (1H, br s, H-15), 2.29 (3H, s, =NCH₃); ^{13}C NMR (CDCl_3) δ 133.0 (C-17), 121.5 (C-19), 54.2, 49.2 (NHCH), 46.0 (=NCH₃).

19-Dimethylamino-8,9-epoxyherbimycin A (9): MS m/z 633 (M, $\text{C}_{32}\text{H}_{47}\text{N}_3\text{O}_{10}$); TLC Rf 0.22 (C_6H_6 - EtOAc, 1:1); $[\alpha]_{\text{D}}^{25} +96^\circ$ (c 0.2, CHCl_3); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ) 268 (19,000) and 325 (1,000); ^1H NMR (CDCl_3) δ 6.37 (1H, d, $J=2.0$ Hz, H-

17), 4.47 (1H, d, $J=2.0$ Hz, H-15), 3.05 (6H, s, $\text{N}(\text{CH}_3)_2$); ^{13}C NMR (CDCl_3) δ 126.4 (C-17), 117.4 (C-19), 67.9 (C-9), 60.1 (C-8), 42.2 ($\text{N}(\text{CH}_3)_2$).

19-Methylpiperazino-8,9-epoxyherbimycin A (10): MS m/z 688 (M, $\text{C}_{35}\text{H}_{52}\text{N}_4\text{O}_{10}$); TLC Rf 0.43 (C_6H_6 - EtOAc, 1:5); $[\alpha]_{\text{D}}^{25} +98^\circ$ (c 0.2, CHCl_3); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ) 261 (23,000); ^1H NMR (CDCl_3) δ 6.42 (1H, d, $J=2.0$ Hz, H-17), 4.50 (1H, d, $J=2.0$ Hz, H-15), 2.33 (1H, s, =NCH₃); ^{13}C NMR (CDCl_3) δ 130.9 (C-17), 119.5 (C-19), 68.8 (C-9), 60.6 (C-8), 56.2, 49.8 (=NCH₂), 46.9 (=NCH₃).

19-Cyclopropylamino-8,9-epoxyherbimycin A (11): MS m/z 645 (M, $\text{C}_{33}\text{H}_{47}\text{N}_3\text{O}_{10}$); TLC Rf 0.15 (C_6H_6 - EtOAc, 1:1); $[\alpha]_{\text{D}}^{25} +135^\circ$ (c 0.3, CHCl_3); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ) 265 (21,500); ^1H NMR (CDCl_3) δ 6.68 (1H, br d, NH), 6.50 (1H, d, $J=1.8$ Hz, H-17), 4.57 (1H, br s, H-15).

19-Dimethylaminoherbimycin A-7,9-cyclic

Table 1. Antitumor activity of herbimycin A derivatives on Ehrlich ascites carcinoma.

Compound	Total dose (mg)	Dose (mg/kg × days)	T/C (%)	No. of cured mice*/total*
1	5	0.5 × 10	109	0/4
2	200	20 × 10	141	1/3
3	500	50 × 10	141	1/3
4	100	11 × 9	122	0/3
5	100	11 × 9	131	0/3
6	100	11 × 9	141	0/3
7	200	22 × 9	110	0/3
8	500	50 × 10	141	2/3
9	500	55.6 × 9	95	0/5
10	750	75 × 10	141	2/3
11	200	22 × 9	114	0/3
12	50	5.5 × 9	110	0/3
13	200	22 × 9	114	0/3

* No. of mice without accumulation of abdominal ascites at day 31.

Table 2. Antitumor activity of 19-methylpiperazino-8,9-epoxyherbimycin A (10).

Tumor	Mouse	Inoculum site	Median survival days of control	Dose (mg/kg × days)	T/C (%)	
Sarcoma 180 (ascites form)	ICR	1 × 10 ⁵	ip	22	100 × 5	200
(solid)	ICR	1 × 10 ⁶	sc	36	12.5 × 5	167
Ehrlich carcinoma	<i>ddY</i>	2.5 × 10 ⁶	ip	17	100 × 5	205
IMC carcinoma	CDF ₁	3 × 10 ⁵	ip	12	200 × 5	358
Meth-A fibrosarcoma	BALB/c	1 × 10 ⁵	ip	29	100 × 5	103
B-16 melanoma	BDF ₁	1 × 10 ⁵	sc	22	25 × 5	155
Lewis lung carcinoma	BDF ₁	1 × 10 ⁵	sc	29	200 × 5	93
P-388 leukemia	CDF ₁	1 × 10 ⁵	ip	12	50 × 5	91
L-1210 leukemia	CDF ₁	1 × 10 ⁵	ip	7	100 × 5	114

Tumors were inoculated sc or ip into mice on day 0. Various doses (12.5~200 mg/kg/day) of an agent were administered ip daily from day 1 to day 5. The observation was terminated at day 60.

carbamate (12): MS *m/z* 633 (M, C₃₂H₄₇N₃O₁₀); TLC Rf 0.71 (EtOAc); [α]_D²⁵ -78° (c 0.1, CHCl₃); UV λ_{max}^{OH} nm (ε) 258 (23,000); ¹H NMR (CDCl₃) δ 6.40 (1H, d, *J*=1.9 Hz, H-17), 4.36 (1H, br s, H-15), 3.16 (6H, s, N(CH₃)₂); ¹³C NMR δ 136.1 (C-17), 115.4 (C-19), 86.4 (C-8), 58.8 (C-9), 43.0 (N(CH₃)₂).

19-Cyclopropylaminoherbimycin A-7,9-cyclic carbamate (13): MS *m/z* 645 (M, C₃₃H₄₇N₃O₁₀); TLC Rf 0.21 (C₆H₆ - EtOAc, 1:1); [α]_D²⁵ -132° (c 0.1, CHCl₃); UV λ_{max}^{OH} nm (ε) 246 (25,000); ¹H NMR (CDCl₃) δ 6.40 (1H, d, *J*=1.8 Hz, H-17), 4.36 (1H, d, *J*=1.8 Hz, H-15), 5.10 (1H, br d, NH).

The introduction of an amino group at the 17-position in compounds 5 and 7 was established

by the appearance of singlet signals of H-15 and H-19 (the signals of both protons of 1 appeared as doublets arising from long-range couplings between H-15 and H-17/H-17 and H-19). On the other hand, a long-range coupling between H-15 and H-17 remained in the 19-amino derivatives (4, 6 and 8~13).

To evaluate the antitumor activity of these derivatives, Ehrlich carcinoma cells (2.5 × 10⁶) were inoculated ip to *ddY* mice on day 0. Mice received various doses of herbimycin A derivatives for 9 or 10 successive days. Antitumor activity was expressed as T/C (%) only at the optimal dose for each derivative: "T" is median survival days of the treated group and "C" is that of the control group. Accumulations of

abdominal ascites were also observed to determine the therapeutic effect on ascites tumor at day 31 when the experiment was terminated.

As shown in Table 1, although herbimycin A (1) did not possess strong antitumor activity, 8,9-epoxyherbimycin A (2), herbimycin A-7,9-cyclic carbamate (3) and the amino derivatives (4, 5, 8 and 10) showed life prolongation of tumor bearing mice. The antitumor activity of 10 against several murine tumors was further examined.

As seen from Table 2, a derivative (10) showed marked antitumor activity against Sarcoma 180 (both ascitic and solid forms), Ehrlich and IMC carcinomas, and B-16 melanoma. The introduction of a methylpiperazino group to the 19 position of the benzoquinone nucleus resulted in high antitumor activity.

References

- 1) ŌMURA, S.; Y. IWAI, Y. TAKAHASHI, N. SADAKANE, A. NAKAGAWA, H. ŌIWA, Y. HASEGAWA & T. IKAI: Herbimycin, a new antibiotic produced by a strain of *Streptomyces*. J. Antibiotics 32: 255~261, 1979
- 2) DUROS, J. & M. SUFFNESS: New antitumor substances of natural origin. In Cancer Treatment Review. 8. pp. 63~87, Academic Press, New York, 1981
- 3) ŌMURA, S.; A. NAKAGAWA & N. SADAKANE: Structure of herbimycin, a new ansamycin antibiotic. Tetrahedron Lett. 1979: 4323~4326, 1979
- 4) FURUSAKI, A.; T. MATSUMOTO, A. NAKAGAWA & S. ŌMURA: Herbimycin A: An ansamycin antibiotic; X-ray crystal structure. J. Antibiotics 33: 781~782, 1980
- 5) DEBOER, C.; P. A. MEULMAN, R. J. WNUK & D. H. PETERSON: Geldanamycin, a new antibiotic. J. Antibiotics 23: 442~447, 1970
- 6) SASAKI, K.; K. L. RINEHART, G. SLOMP, M. F. GROSTIC & E. C. OLSON: Geldanamycin. I. Structure assignment. J. Am. Chem. Soc. 92: 7591~7593, 1970
- 7) TANIDA, S.; T. HASEGAWA & E. HIGASHIDE: Macbecins I and II, new antitumor antibiotics. I. Producing organism, fermentation and antimicrobial activities. J. Antibiotics 33: 199~204, 1980
- 8) MUROI, M.; M. IZAWA, Y. KOSAI & M. ASAI: Macbecins I and II, new antitumor antibiotics. II. Isolation and characterization. J. Antibiotics 33: 205~212, 1980
- 9) MUROI, M.; K. HAIBARA, M. ASAI & T. KISHI: The structures of macbecins I and II, new antitumor antibiotics. Tetrahedron Lett. 1980: 309~312, 1980
- 10) SASAKI, K.; H. YASUDA & K. ONODERA: Growth inhibition of virus transformed cells *in vitro* and antitumor activity *in vivo* of geldanamycin and its derivatives. J. Antibiotics 32: 849~851, 1979